
Antiproliferative properties of iron supramolecular cylinders

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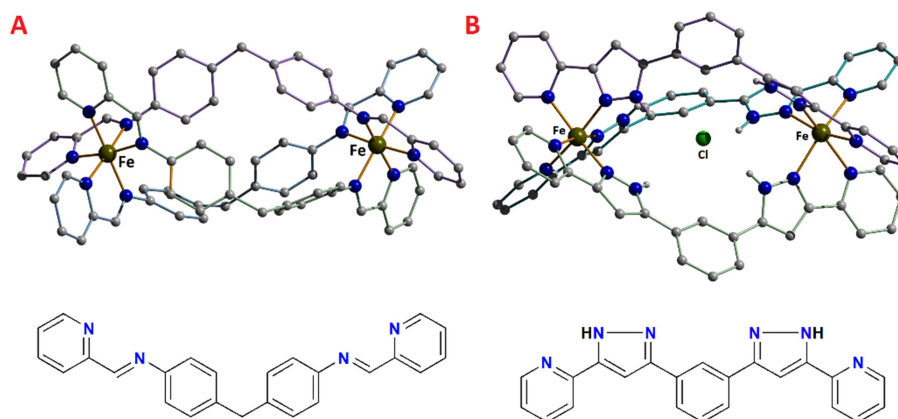


Figure S1. Representation of the crystal structures (cationic part) of known iron-based helicates together with their corresponding ligand, which were used in the present study for comparison purposes. A) $[\text{Fe}_2(\text{C}_{25}\text{H}_{20}\text{N}_4)_3]\text{Cl}_4$ (**R1**) [1] and B) $\text{Cl}@[Fe_2(\text{C}_{22}\text{H}_{16}\text{N}_6)_3]\text{Cl}(\text{PF}_6)_2$ (**R2**) [2].

Table S1. Crystal data and structure refinement for metallohelicites **H1** and **H2**.

Compound	H1	H2
Empirical formula	$\text{C}_{99}\text{H}_{65}\text{Fe}_2\text{NO}_{12.25}$	$\text{C}_{76}\text{H}_{58}\text{Fe}_2\text{N}_2\text{O}_{20}$
Formula weight (g mol^{-1})	1576.22	1430.94
Temperature (K)	100(2)	100(2)
Crystal system	triclinic	monoclinic
Space group	$P\bar{1}$	$C2/c$
Crystal size (mm^3)	$0.20 \times 0.04 \times 0.03$	$0.06 \times 0.02 \times 0.01$
a (Å)	15.300(2)	23.016(4)
b (Å)	16.940(3)	17.915(3)
c (Å)	17.496(3)	18.590(6)
α (°)	64.285(2)	90
β (°)	72.373(2)	121.900(2)
γ (°)	82.106(2)	90
V (Å ³)	3893.6(11)	6508(3)
Z	2	4
ρ_{calcd}	1.344	1.461
μ (mm^{-1})	0.551	0.661
$F(000)$	1632	2960
θ for data collection (°)	2.85– 26.22	2.71– 33.17
Reflections collected / unique	56858 / 11975	32347 / 8404
Completeness to theta	0.991	0.998
Data / restraints / parameters	11975 / 586 / 1161	8404 / 112 / 495
Goodness-of-fit on F^2	1.074	1.034
Final R indices [$I > 2\sigma(I)$]	$R1 = 0.0815$, $wR2 = 0.1960$	$R1 = 0.0601$, $wR2 = 0.1524$
R indices (all data)	$R1 = 0.1263$, $wR2 = 0.2245$	$R1 = 0.1022$, $wR2 = 0.1743$
largest diff. peak and hole ($e \text{ Å}^{-3}$)	0.713 and -0.407	1.162 and -0.570

Table S2. Selected bond lengths (Å) and angles (°) for metallohelicates **H1** and **H2**.

H1			
Fe1–O1	2.018(5)	Fe1–O2	2.001(5)
Fe1–O5	1.975(4)	Fe1–O6	2.000(4)
Fe1–O9	1.980(4)	Fe1–O10	1.966(4)
Fe2–O3	1.977(5)	Fe2–O4	1.987(5)
Fe2–O7	1.977(5)	Fe2–O8	1.979(6)
Fe2–O11	1.982(4)	Fe2–O12	1.970(5)
O1–Fe1–O2	85.64(19)	O2–Fe1–O6	83.60(18)
O6–Fe1–O9	97.38(18)	O9–Fe1–O1	94.25(18)
O2–Fe1–O5	97.48(18)	O5–Fe1–O9	88.63(18)
O9–Fe1–O10	86.05(17)	O10–Fe1–O2	87.93(17)
O3–Fe2–O4	87.0(2)	O4–Fe2–O8	87.6(2)
O8–Fe2–O11	97.1(2)	O11–Fe2–O3	88.69(18)
O4–Fe2–O7	95.80(19)	O7–Fe2–O11	86.95(17)
O11–Fe2–O12	86.88(18)	O12–Fe2–O4	90.7(2)
H2			
Fe1–O2	1.969(2)	Fe1–O3	1.966(3)
Fe1–O8	2.010(2)	Fe1–O9	1.973(2)
Fe1–O4a ^a	2.021(2)	Fe1–O5a ^a	2.008(3)
O2–Fe1–O3	86.75(10)	O3–Fe1–O9	87.30(10)
O9–Fe1–O5a	100.21(11)	O5a–Fe1–O2	86.64(10)
O3–Fe1–O4a	86.81(10)	O4a–Fe1–O5a	84.97(10)
O5a–Fe1–O8	91.41(11)	O8–Fe1–O3	97.90(10)

^aSymmetry operation: a = 1–x, y, 1/2–z.**Table S3.** Selected contact distances (Å) for metallohelicates **H1** and **H2**.

H1		π – π interactions
C58B···C64Bf	3.383(18)	
centroid···centroid	3.712(2)	
H2		Hydrogen bonds and π – π interactions
O1–H1···O4	2.748(4)	165(5)
O6a ^a –H6a···O1w	2.611(15)	164
O7–H7···N1s	2.756(6)	152(4)
C1···C7	3.326(8)	

^aSymmetry operation: a = 1–x, y, 1/2–z, 1+z; f = 1–x, –y, 2–z

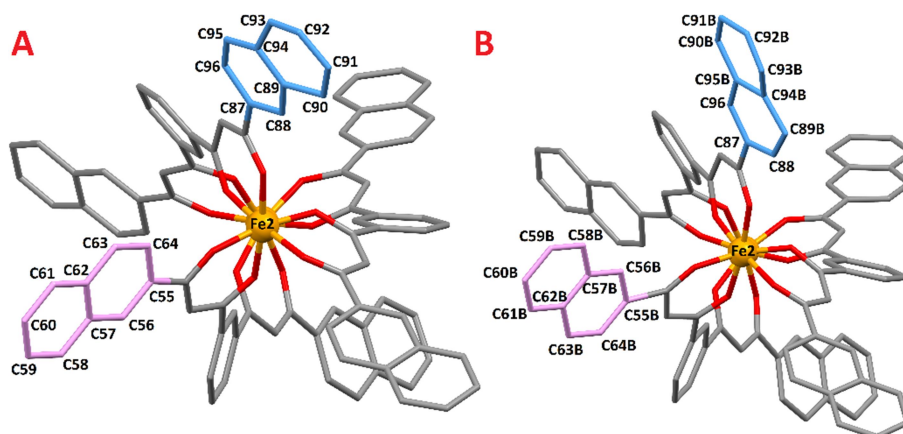


Figure S2. Major (60%) and minor (40%) conformations of **H1** present in the crystal lattice.

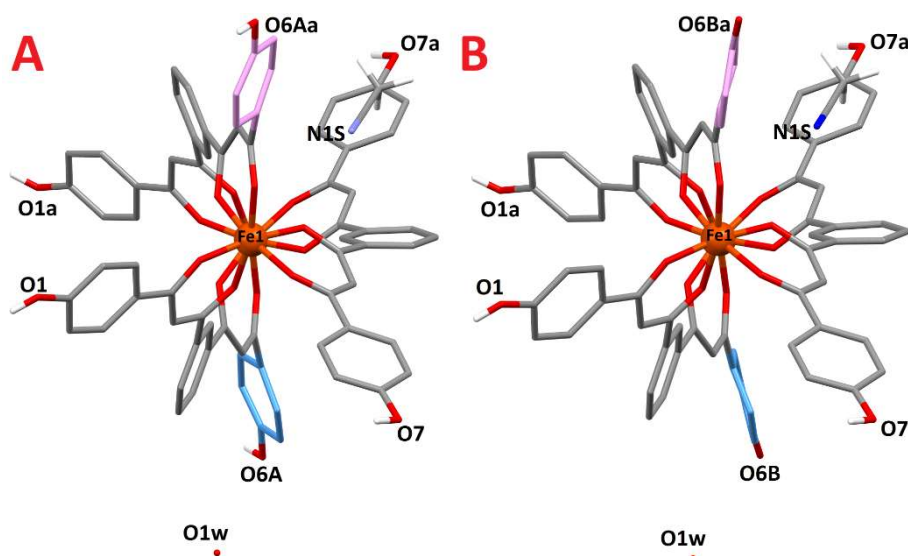


Figure S3. Major (60%) and minor (40%) conformations of **H2** present in the crystal lattice.

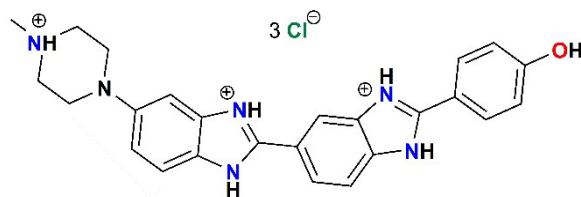


Figure S4. Representation of Hoechst 33258 (2'-(4-hydroxyphenyl)-5-(4-methyl-1-piperazinyl)-2,5'-bi-1*H*-benzimidazole trihydrochloride hydrate).

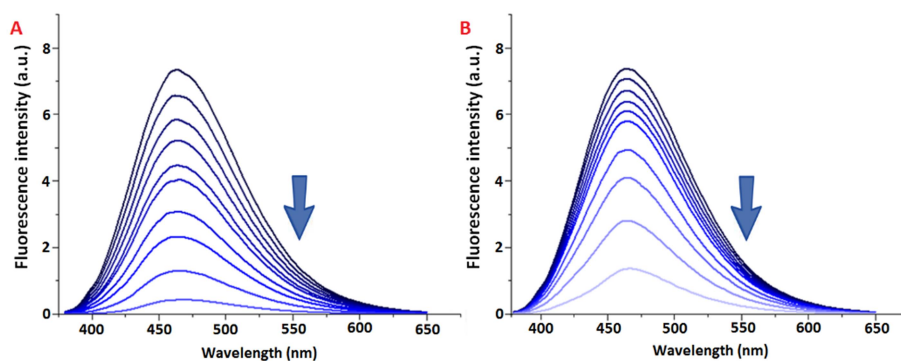


Figure S5. Emission spectra of DNA-Hoechst complex ([DNA] = 0.19 μ M and [Hoechst 33258] = 15 μ M), λ_{exc} = 349 nm, λ_{em} = 458 nm, upon addition of increasing amounts (2–150 μ M) of A) **H1** and B) **H2**. The blue arrows show the diminution of the emission intensity with the increase of the complex concentration.

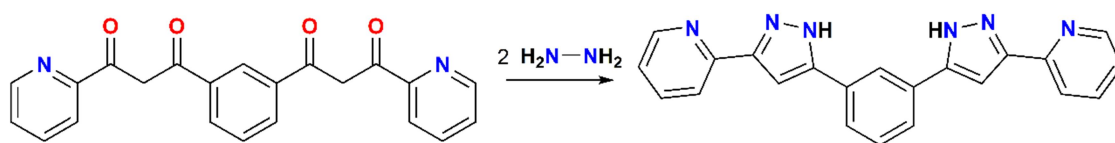


Figure S6. Preparation of the ligand found in $\text{Cl}@\text{[Fe}_2(\text{C}_{22}\text{H}_{16}\text{N}_6)_3\text{]Cl(PF}_6)_2$ (**R2**), which is obtained from a bis- β -diketone and hydrazine [2].

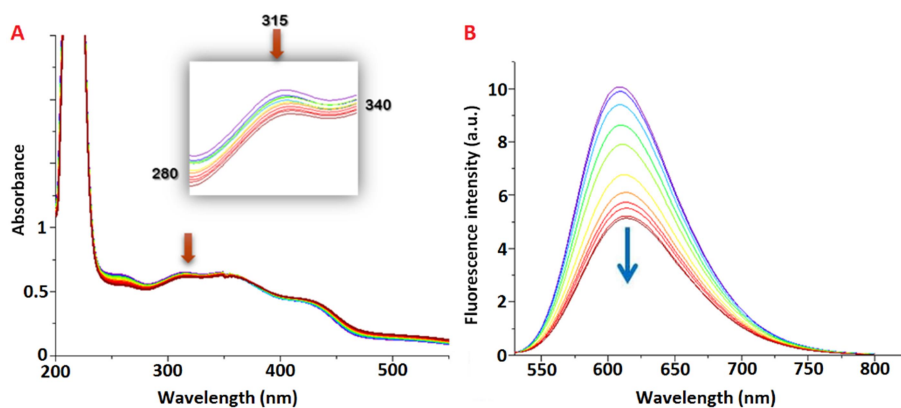


Figure S7. A) Absorption spectra of **R2** in 10 mM sodium cacodylate/20 mM NaCl buffer upon addition of ct-DNA (0–25 μM). The inset shows a zoom of the absorption region (MLCT band) used to determine the intrinsic binding constants K_b . B) Emission spectra of DNA-EB (obtained using [ct-DNA] = 25 μM and [EB] = 125 μM) in 10 mM sodium cacodylate, 20 mM NaCl, λ_{exc} = 514 nm, λ_{em} = 610 nm, upon addition of increasing amounts of **R2** (2.5–50 μM). The blue arrow shows the diminution of the emission intensity with the increase of complex concentration. The [ct-DNA]_{bp} was determined from its absorption intensity at 260 nm, with a molar extinction coefficient of 6600 M⁻¹cm⁻¹ [3].

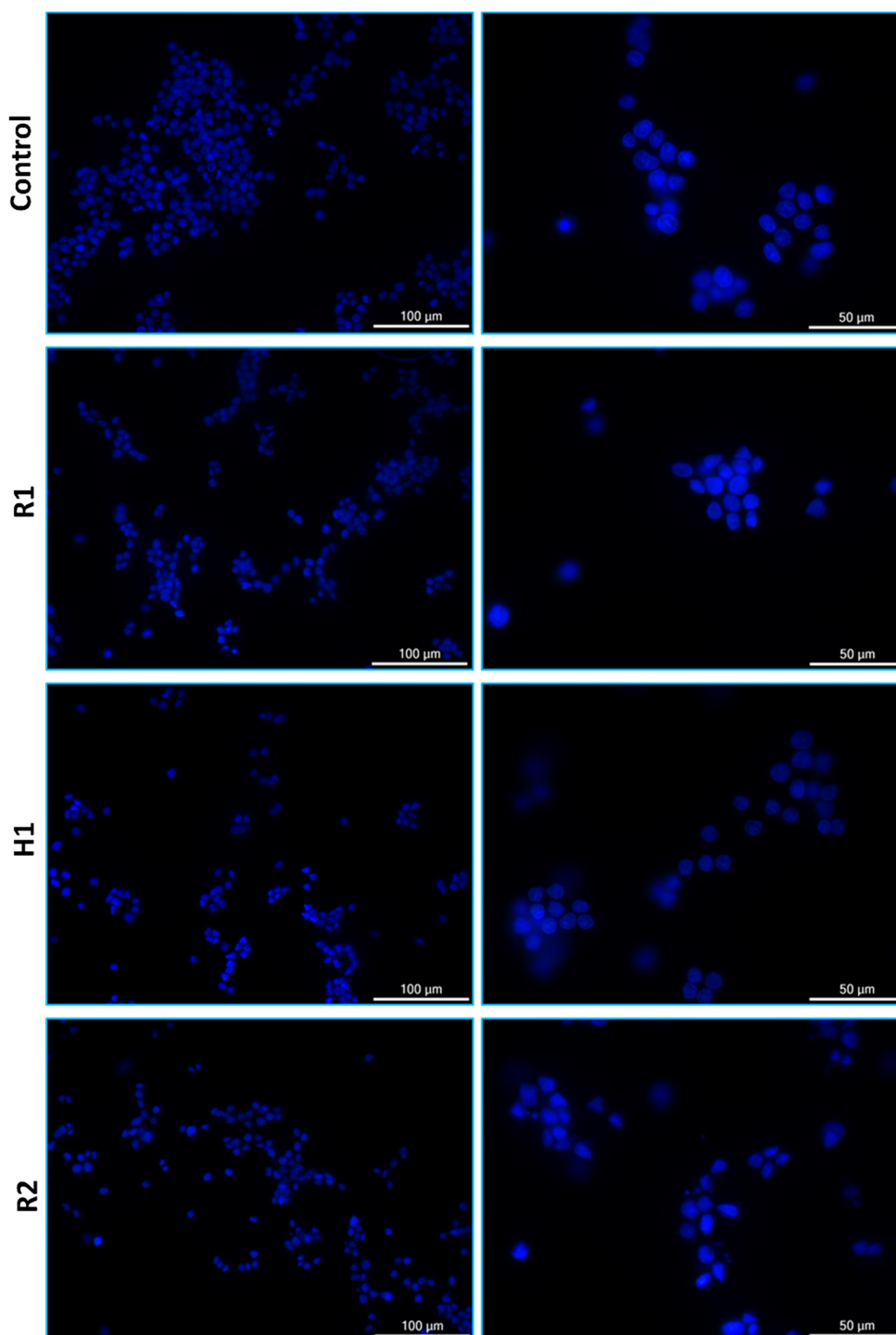


Figure S8. Confocal microscopy images showing the blue-stained nucleus of SW620 cells, SW620 cells incubated with **R1**, SW620 cells incubated with **H1** and SW620 cells incubated with **R2**. The concentrations of each compound used correspond to the corresponding IC₅₀ values (see Table 2). The selected images are representative of three distinct series of images. Incubation time = 48 h; scale bars: left images, 100 μm and right images, 50 μm .

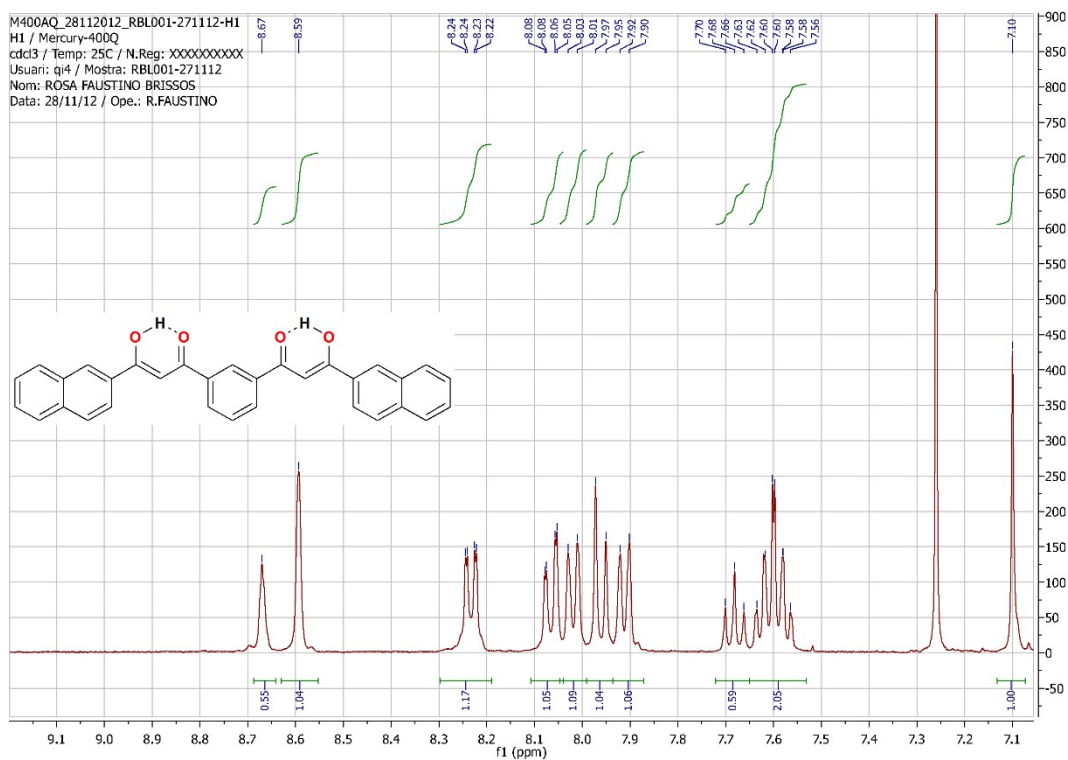


Figure S9. ¹H NMR spectrum of ligand **H₂L1** in CDCl₃. The enolic O–H proton is not seen (usually such acidic protons appear at $\delta > 15$ ppm).

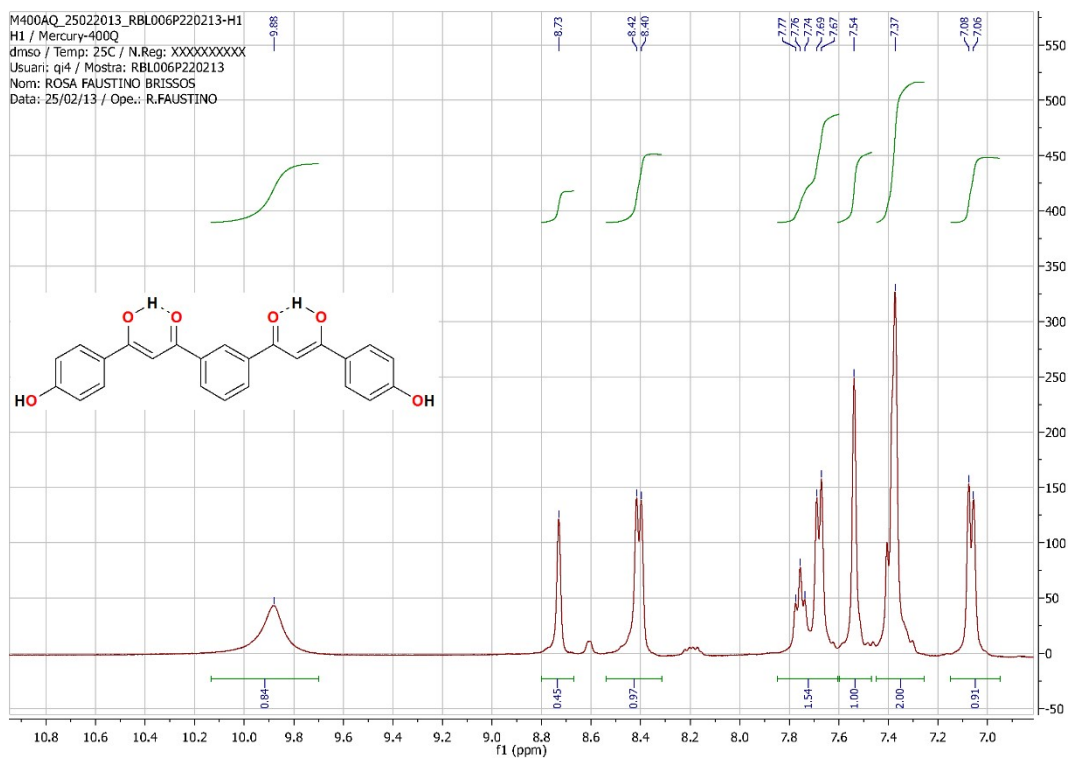
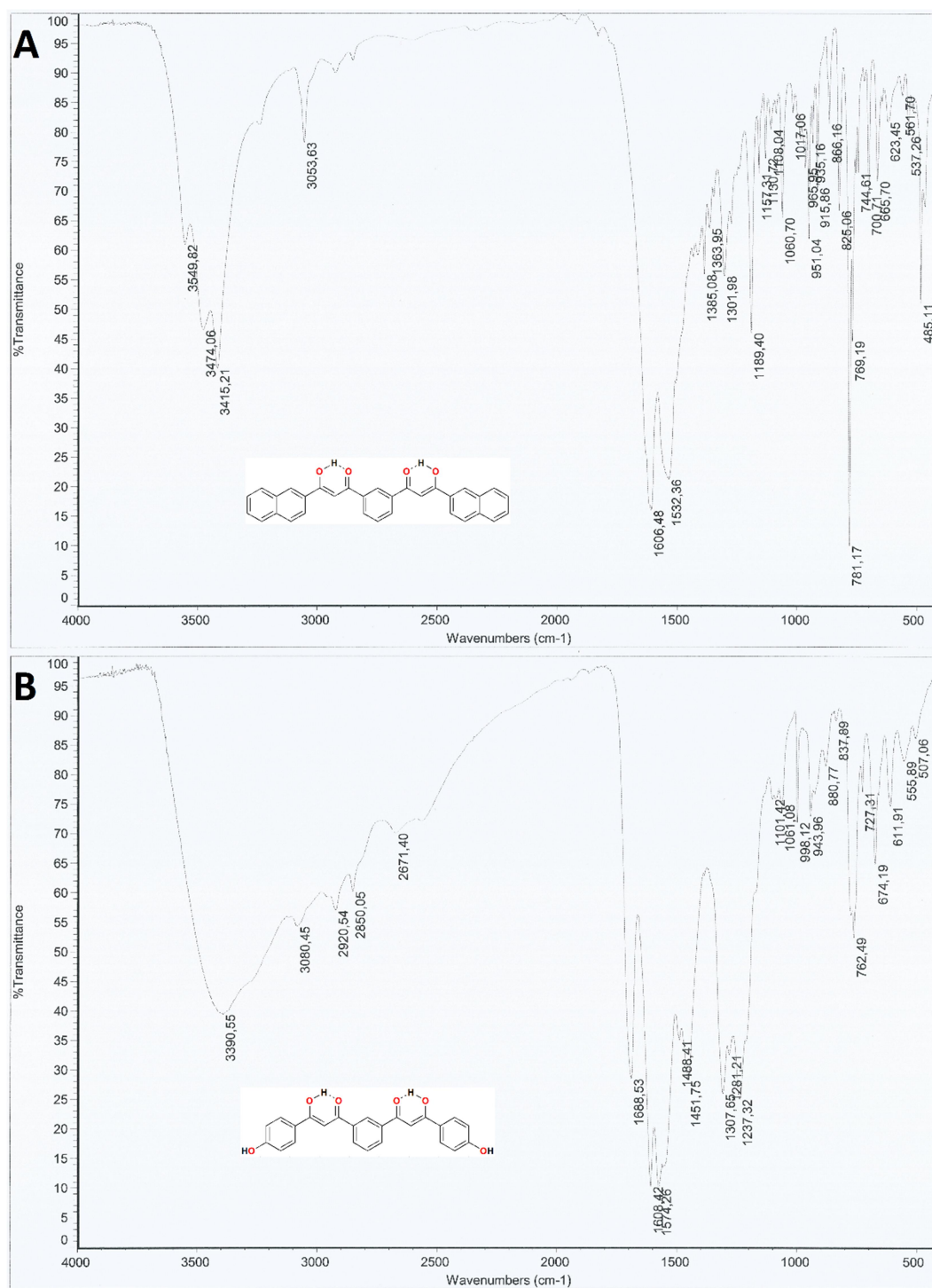


Figure S10. ¹H NMR spectrum of ligand **H₄L2** in d₆-DMSO. The enolic O–H proton is not seen (usually such acidic protons appear at $\delta > 15$ ppm).

Figure S11. IR spectrum of A) ligand **H₂L1** and of B) ligand **H₄L2**.

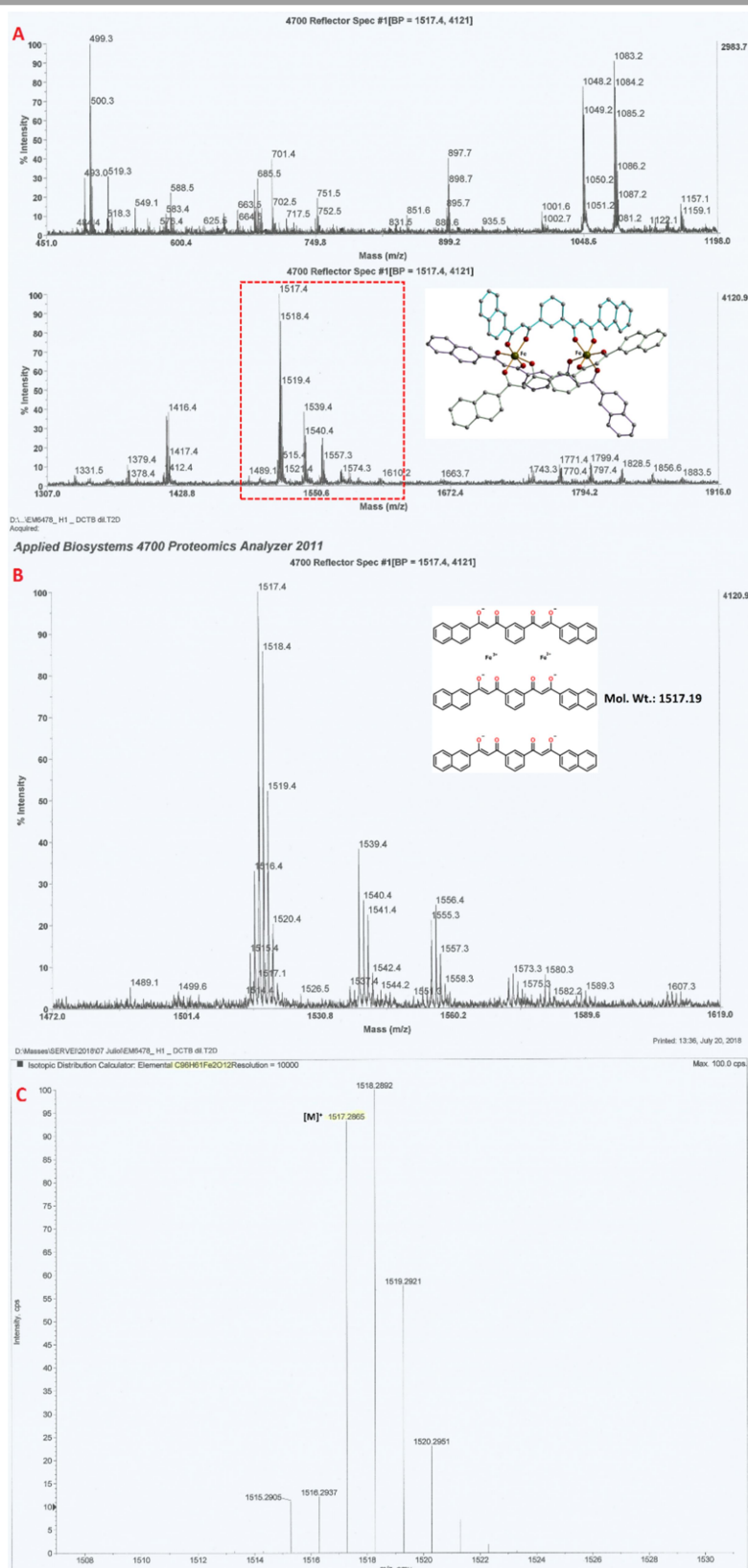


Figure S12. A) MALDI-TOF spectrum of **H1**; B) zoom of the Mol. Wt. region of the spectrum; C) theoretical isotopic pattern for the molecular peak of **H1**.

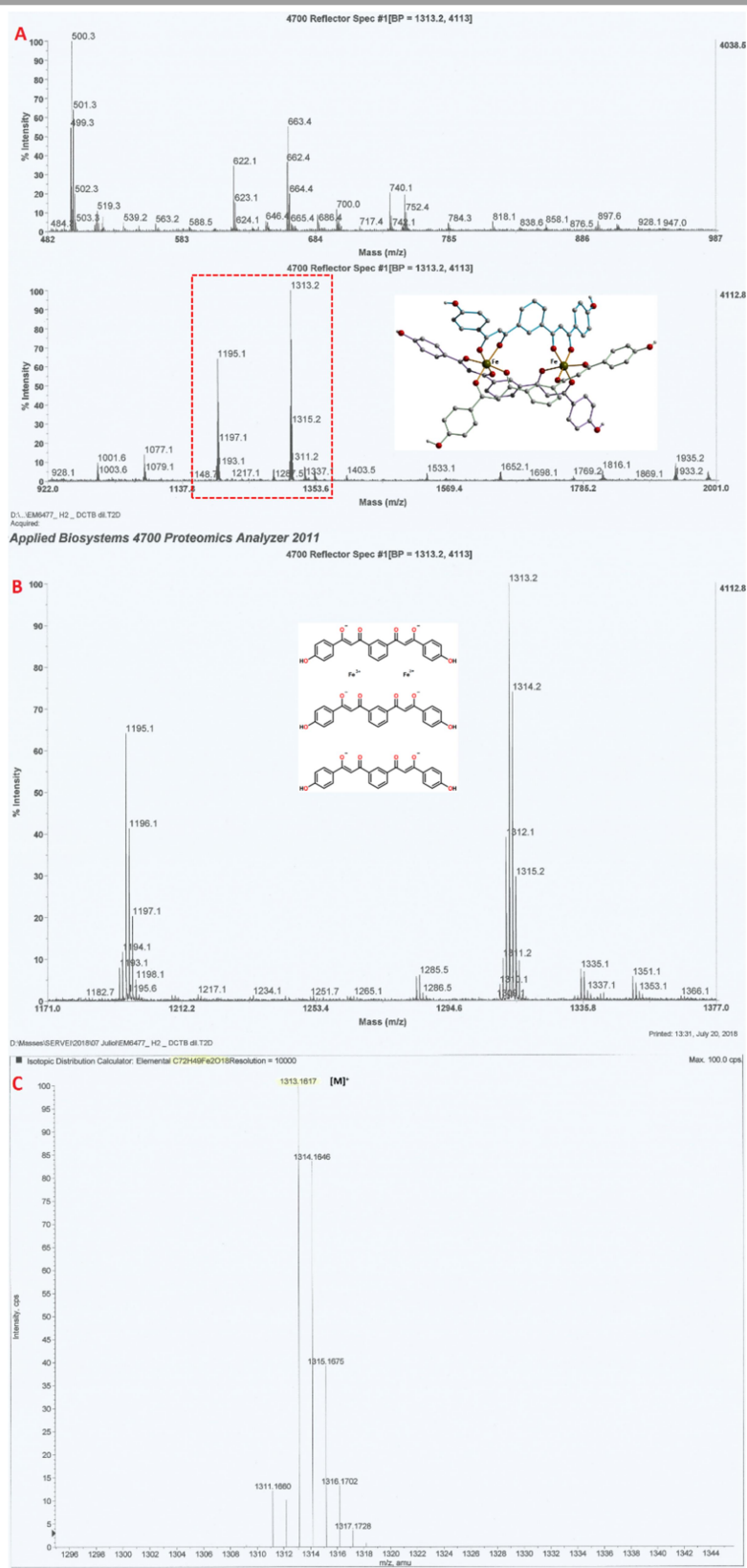


Figure S13. A) MALDI-TOF spectrum of **H2**; B) zoom of the Mol. Wt. region of the spectrum; C) theoretical isotopic pattern for the molecular peak of **H2**.

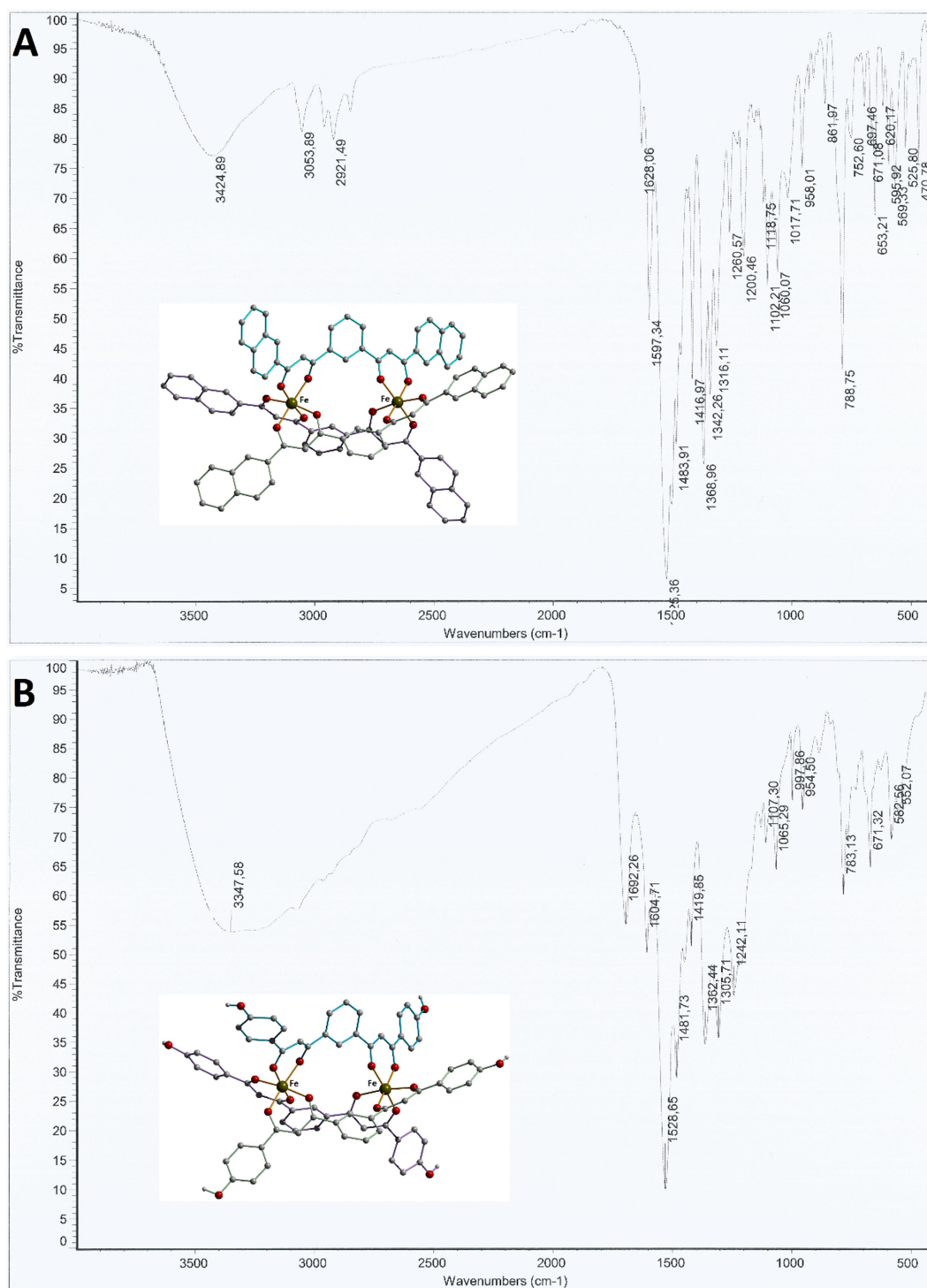


Figure S14. IR spectrum of A) helicate **H1** and of B) helicate **H2**.

References

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